Identification, astaxanthin production and genome sequence of yeasts from flowers

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Astaxanthin is a natural carotenoid pigment with high antioxidant activity and commercial value. Microbial astaxanthin production has become a promising alternative. A total of 29 yeasts were isolated from 50 flower samples collected in Lampang province, Thailand. Of these, 17 isolates were determined to be Basidiomycota, belonging to Rhodotorula paludigena (11 strains), R. mucilaginosa (3 strains), and potential new species closely related to R. kratochvilovae (3 strains). Meanwhile, the 12 remaining isolates were classified as 9 species in 7 genera of Ascomycota. Surprisingly, all the basidiomycetous yeasts found in this study were pink to orange-red pigmented and showed positive astaxanthin formation when determined on a TLC-developed plate. Analysis of their intracellular astaxanthin accumulation by UV spectrometry revealed that R. paludigena TL35-5 was the greatest astaxanthin-producing strain with an astaxanthin content of 275.94 ± 0.16 g/g DCW. R. paludigena was first reported as astaxanthin-producing yeasts in this study. R. paludigena TL35-5 was then selected for further astaxanthin production optimization and genome sequence analysis. The whole genome of *R. paludigena* is the second to be reported to date for its species. R. paludigena TL35-5 genome was 20,982,417 bp long with a GC content of 63.75%. A total of 6,701 protein-encoding genes were predicted, with a total length of 10,181,859 bp. Essential genes required for the biosynthesis of β-carotene, and other derivative biosynthesis pathways namely terpenoid backbone biosynthesis, xanthophyll biosynthesis, and β-carotene degradation, could be identified. Our findings indicated that *R. paludigena* TL35-5 had the potential to be an astaxanthin-producing yeast.